Scaling of xylem vessels and veins within the leaves of oak species

David A. Coomes1,*, Steven Heathcote1, Elinor R. Godfrey1, James J. Shepherd1, and Lawren Sack2

1University of Cambridge, Downing Street, Cambridge CB2 3EA, UK
2Department of Ecology and Evolutionary Biology, University of California, Los Angeles, 621 Charles E. Young Drive South, Los Angeles, CA 90095-1606, USA
*Author for correspondence (dac18@cam.ac.uk).

General models of plant vascular architecture, based on scaling of pipe diameters to remove the length dependence of hydraulic resistance within the xylem, have attracted strong interest. However, these models have neglected to consider the leaf, an important hydraulic component; they assume all leaves to have similar hydraulic properties, including similar pipe diameters in the petiole. We examine the scaling of the leaf xylem in 10 temperate oak species, an important hydraulic component. The mean hydraulic diameter of petiole xylem vessels varied by 30% among the 10 oak species. Conduit diameters narrowed from the petiole to the midrib to the secondary veins, consistent with resistance minimization, but the power function scaling exponent differed from that predicted for stems. Leaf size was an organizing trait within and across species. These findings indicate that leaf vasculature needs to be included in whole-plant scaling models, for these to accurately reflect and predict whole-plant transport and its implications for performance and ecology.

Keywords: tapering; xylem vessels; hydraulic resistance; scaling relationships; allometry; stomatal density

1. INTRODUCTION

Plants need to deliver copious water to the leaf cells and airspaces via narrow conduits. If the entire vascular system comprised narrow conduits, its hydraulic resistance would be very high because conduit lumen resistance scales inversely with diameter raised to the fourth power (the Hagen–Poiseuille law; Tyree & Zimmermann 2002). Such a high hydraulic resistance would be very high because conduit lumen diameter (dh) tapers with branch diameter (D) as dh ∝ Dα (where optimal α = 1/6; Anfodillo et al. 2006; Coomes et al. 2006). In fact, the leaf xylem may follow different rules given its different structure and function, e.g. its reticulation and leakiness to the mesophyll surrounding the veins (McCulloh & Sperry 2005a,b).

We examined scaling relationships within the leaf vasculature of 10 temperate oak species (Quercus, Fagaceae). For a given species, and across species, we hypothesized that there would be tapering of dh in petiole, midrib and secondary veins, and of vascular bundles within veins. Further, we hypothesized that the dh would increase with leaf size within and across species. Additionally, we tested whether the leaf xylem vessel properties correlate across species with key functional traits, stomatal density (SD) and leaf mass per unit area (LMA).

2. MATERIAL AND METHODS

Leaves from 10 oak species were sampled from the University Botanic Garden, Cambridge, UK and the Sir Harold Hillier Gardens, Hampshire, UK. Most of these trees were planted within the last 30 years and their diameters at breast height ranged from 1.2 to 141 cm (mean 40 cm). We sampled disease-free leaves from a sun-exposed, south-facing branch for five trees per species. Lamina areas were determined from digital images using IJ (http://rsb.info.nih.gov/ij/). We determined the SD using nail varnish imprints of the abaxial leaf surfaces; we counted stomata within three areas of the peel under an optical microscope (×100; L3000B-HTG, GX Optical, China). LMA was determined by dividing leaf area by mass after drying the leaves for 72 hours at 70°C.

We estimated the diameters of the vascular bundles and vessels from cross-sections at (i) the midpoint of the petiole, (ii) one-third of the way along the midrib, and (iii) one-third of the way along the secondary vein branching closest to the point at which the midrib was cut. We stained the sections with Safranin O (0.01%) for imaging and measurement of diameters of vessel lumens and vascular bundles (including fibres) using a confocal microscope with an Argon laser and analysis software (LCSPP2 model, Leica Microsystems, Germany). The hydraulic conductance of a pipe scales as the fourth power of lumen diameter (dh), so we calculated a hydraulically weighted mean diameter, dh = ∑ dhj/∑ dhj (Mencuccini et al. 1997; Anfodillo et al. 2006), using all vessels with a diameter greater than half that of the largest vessel in each sample to eliminate vessels that appear to be narrow because the tapering ends have been sampled (James et al. 2003), and the number of such vessels within a bundle was counted (N).

We determined the scaling of dh and the vascular bundle diameter D by fitting straight lines to log-log transformed data (i.e. allometric relationships) by using the standard major axis regression (SMA; using the SMATR package in the R statistics software); this also allowed a comparison of whether slopes (α values) and intercepts differed among species. The SMA

Published online 11 April 2008

Received 19 February 2008
Accepted 19 March 2008

This journal is © 2008 The Royal Society
It is now widely acknowledged that the narrowing in petioles, midribs and secondary veins (figure A1 in the electronic supplementary material). Within and across the 10 species, there were strong scaling relationships between $d_h$, and $D$ (figures 1 and 2). Within and across species, $d_h$, correlated with $D$ for petioles, midrib and secondary veins (figure A1 in the electronic supplementary material, figure 1a–c); additionally, the $D$ of petioles, midribs and secondary veins were intercorrelated (figure 1d–f), as were the $d_h$ in petioles, midribs and secondary veins (figure 1g–i). Across vein orders, for all the 10 species, $d_h$, scaled with $D$, with $\alpha$ values of 0.42–0.73, substantially greater than the 1/6 predicted by scaling models for stems (figure 1; table 1).

Across species, $d_h$, and $D$, and petioles, midribs and secondary veins scaled tightly with mean leaf area (figure 2). The species’ $\alpha$ values correlated negatively with the leaf area, indicating a steeper conduit tapering in smaller leaves (table 1; Spearman’s rank test, $p = 0.032$). The leaf xylem properties were not correlated with LMA, SD or the average size of the adult tree (figure 2).

### 4. DISCUSSION

It is now widely acknowledged that the narrowing of conduit diameters towards the periphery of bulk transport systems reduces the dependency of hydraulic resistance on path length (Becker et al. 2000), but controversy remains over which processes need to be included in models in order to accurately predict the scaling of $d_h$, and $D$. The pioneering model of West et al. (1999) predicts that the length dependence of hydraulic resistance is practically removed from branching stem systems when $d_h \propto D^{1/6}$, assuming that the system comprises continuous non-leaky pipes running from trunk to the petioles and that branching systems are fractal-like and volume filling (West et al. 1999; Becker et al. 2000; Anfodillo et al. 2006). We have shown strong relationships between $d_h$, and $D$ for the leaf vasculature of the 10 oak species with scaling exponents consistently greater than 1/6. Several factors will need to be considered in the derivation of a theoretical model of vascular architecture for leaves as distinct from that of stems. Leaf venation is approximately plane filling and typically reticulate (Pelletier & Turcotte 2000). Also, leaf venation is laterally ‘leaky’ to the surroundings (McCulloh & Sperry 2005a,b; Zwieniecki et al. 2006); the tapering of vascular bundles within pine needles is optimized to enable even delivery of water along their length (Zwieniecki et al. 2006). In addition, the cost of constructing the vascular system needs to be considered to determine the theoretically optimal scaling of $d_h$, with $D$ (McCulloh & Sperry 2005a,b). Further investigation is needed of the developmental, hydraulic and economic factors that would determine the optimal tapering of leaf xylem conduits.

Our observations of general scaling between leaf size and vascular properties are consistent with early demonstrations that the leaf area correlates with the petiole vessel diameter for a given species (Salisbury 1913). This scaling supports the classical principle that larger leaves would require a larger ‘pipe’ system to supply a greater transpiration surface (cf. Shinozaki et al. 1964). We also found that the scaling of $d_h$, with $D$ itself scaled with the leaf size across species. More work is needed to determine how far phylogenetically diverse species depart from the scaling trends found for the 10 oak species. The lack of coordination between $d_h$, and $D$ and SD or LMA needs to be confirmed across diverse species sets.

The $d_h$, values given here are within the range reported previously for other tree species (Cochard et al. 2004; Nardini et al. 2005; Sack & Frede 2006), for which the leaf hydraulic conductance was low, 2–20 mmol m$^{-2}$ s$^{-1}$ MPa$^{-1}$, thus accounting for a major portion of whole-plant resistance. Because the leaf xylem scales differently from tree stem xylem, and species differ in the leaf xylem properties

---

### Table 1. Petiole xylem characteristics of the 10 temperate oak species, including hydraulically weighted mean vessel diameter ($d_h$), vascular bundle diameter ($D$) and number of vessels ($N$), as well as the mean individual leaf area (area), LMA and SD.

(The final column contains the scaling exponent of the power functions shown in figure 1. All of these properties varied significantly among species (ANOVA, $p < 0.05$). n.d., not determined because leaves were hirsute.)

<table>
<thead>
<tr>
<th>Code</th>
<th>Species</th>
<th>$d_h$ (µm)</th>
<th>$N$ (no. mm$^{-2}$)</th>
<th>$D$ (µm)</th>
<th>Area (cm$^2$)</th>
<th>LMA (g m$^{-2}$)</th>
<th>SD (no. mm$^{-2}$)</th>
<th>$\alpha$ values</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td><em>Q. castaneifolia</em></td>
<td>27.4</td>
<td>25.6</td>
<td>968</td>
<td>56</td>
<td>105</td>
<td>332</td>
<td>0.55</td>
</tr>
<tr>
<td>2</td>
<td><em>Q. cerris</em></td>
<td>25.5</td>
<td>51.8</td>
<td>711</td>
<td>26</td>
<td>87</td>
<td>515</td>
<td>0.59</td>
</tr>
<tr>
<td>3</td>
<td><em>Q. georgiana</em></td>
<td>22.4</td>
<td>51.5</td>
<td>670</td>
<td>22</td>
<td>74</td>
<td>752</td>
<td>0.58</td>
</tr>
<tr>
<td>4</td>
<td><em>Q. ilex</em></td>
<td>21.2</td>
<td>21.2</td>
<td>705</td>
<td>22</td>
<td>104</td>
<td>n.d.</td>
<td>0.73</td>
</tr>
<tr>
<td>5</td>
<td><em>Q. nigra</em></td>
<td>26.1</td>
<td>19.2</td>
<td>886</td>
<td>30</td>
<td>74</td>
<td>525</td>
<td>0.48</td>
</tr>
<tr>
<td>6</td>
<td><em>Q. petraea</em></td>
<td>28.2</td>
<td>27.4</td>
<td>1217</td>
<td>85</td>
<td>49</td>
<td>228</td>
<td>0.42</td>
</tr>
<tr>
<td>7</td>
<td><em>Q. rhyphophyla</em></td>
<td>26.8</td>
<td>31.2</td>
<td>1505</td>
<td>74</td>
<td>110</td>
<td>761</td>
<td>0.47</td>
</tr>
<tr>
<td>8</td>
<td><em>Q. robur</em></td>
<td>25.5</td>
<td>30.0</td>
<td>1222</td>
<td>61</td>
<td>54</td>
<td>275</td>
<td>0.48</td>
</tr>
<tr>
<td>9</td>
<td><em>Q. rubra</em></td>
<td>34.0</td>
<td>30.4</td>
<td>1132</td>
<td>128</td>
<td>81</td>
<td>493</td>
<td>0.51</td>
</tr>
<tr>
<td>10</td>
<td><em>Q. serrata</em></td>
<td>30.6</td>
<td>36.6</td>
<td>847</td>
<td>59</td>
<td>74</td>
<td>653</td>
<td>0.60</td>
</tr>
</tbody>
</table>

Regression was also used to fit lines to non-transformed data (i.e. isometric relationships).
sufficiently to influence their whole-plant hydraulic resistance, realistic models of whole-plant vasculature should explicitly include the scaling of leaf venation architecture.

We are grateful to Allen Coombes and the Sir Harold Hillier Gardens. This work was supported by University of Cambridge UROP studentships and by National Science Foundation grant no. IOB-0546784.


Becker, P., Gribben, R. J. & Lim, C. M. 2000 Tapered conduits can buffer hydraulic conductance from path-length effects. Tree Physiol. 20, 965–967.

Brodribb, T. J., Holbrook, N. M., Zwieniecki, M. A. & Palma, B. 2005 Leaf hydraulic capacity in ferns, conifers

Figure 1. Scaling of hydraulically weighted vessel diameter \( (d_h) \) with vascular bundle diameter \( (D) \) within 10 species of oak; standard major axes were fitted to the data. Vessels were sampled from the following: open circles, primary vein; triangles, secondary vein; and filled circles, petiole. (a) Quercus castaneifolia: \( d_h = 0.67D^{0.55} \) and \( r = 0.87, p < 0.001 \); (b) Q. cerris: \( d_h = 0.52D^{0.59} \) and \( r = 0.93, p < 0.001 \); (c) Q. georgiana: \( d_h = 0.53D^{0.58} \) and \( r = 0.68, p < 0.001 \); (d) Q. ilex: \( d_h = 0.18D^{0.72} \) and \( r = 0.85, p < 0.001 \); (e) Q. nigra: \( d_h = 1.07D^{0.48} \) and \( r = 0.78, p < 0.001 \); (f) Q. petraea: \( d_h = 1.61D^{0.42} \) and \( r = 0.61, p < 0.001 \); (g) Q. rhysophylla: \( d_h = 0.95D^{0.47} \) and \( r = 0.67, p < 0.001 \); (h) Q. robur: \( d_h = 1.02D^{0.47} \) and \( r = 0.58, p = 0.001 \); (i) Q. rubra: \( d_h = 1.05D^{0.51} \) and \( r = 0.81, p < 0.001 \); and (j) Q. serrata: \( d_h = 0.52D^{0.60} \) and \( r = 0.98, p < 0.001 \).
Figure 2. Correlations among foliar traits measured for 10 species of oak. The upper right panels contain Pearson correlation coefficients (upper number, all values in bold are greater than 0.60 and significant at $p < 0.05$) and SMA slopes (lower number); none of the variables were transformed. The lower left panels show the data points. The traits are abbreviated along the diagonal as follows: P.D, petiole vein diameter; M.D, midrib vein diameter; S.D, secondary vein diameter; P.dh, petiole vessel mean hydraulically weighted diameter; M.dh, midrib vessel mean hydraulically weighted diameter; S.dh, secondary mean hydraulically weighted vessel diameter; LA, leaf area; LMA, leaf mass per unit area; S, stomatal density.


